

#7 attachment 09/85



D1

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION  
International Bureau

## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>6</sup> :</b> <b>C12N 15/16, C07K 14/65, A61K 38/30</b>	<b>A1</b>	<b>(11) International Publication Number:</b> <b>WO 97/33997</b> <b>(43) International Publication Date:</b> 18 September 1997 (18.09.97)
<b>(21) International Application Number:</b> PCT/GB97/00658 <b>(22) International Filing Date:</b> 11 March 1997 (11.03.97)  <b>(30) Priority Data:</b> 9605124.8                      11 March 1996 (11.03.96)                      GB  <b>(71) Applicant (for all designated States except US):</b> ROYAL FREE HOSPITAL SCHOOL OF MEDICINE [GB/GB]; University of London, Rowland Hill Street, London NW3 2PF (GB).  <b>(72) Inventor; and</b> <b>(75) Inventor/Applicant (for US only):</b> GOLDSPINK, Geoffrey [GB/GB]; Royal Free Hospital School of Medicine, Dept. of Anatomy and Developmental Biology, Division of Basic Medical Sciences, Rowland Hill Street, London NW3 2PF (GB).  <b>(74) Agents:</b> CRESSWELL, Thomas, Anthony et al.; J.A. Kemp & Co., 14 South Square, Gray's Inn, London WC1R 5LX (GB).		<b>(81) Designated States:</b> JP, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).  <b>Published</b> <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> METHOD OF TREATING MUSCULAR DISORDERS  <b>(57) Abstract</b>  An IGF-I polypeptide or functional derivative thereof characterized by the presence of the Ec peptide or functional equivalent thereof for use in a method of treatment or therapy of the human or animal body.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
AU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Ireland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgyzstan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic of Korea	SD	Sudan
CF	Central African Republic	KR	Republic of Korea	SE	Sweden
CG	Congo	KZ	Kazakhstan	SG	Singapore
CH	Switzerland	LI	Liechtenstein	SI	Slovenia
CI	Côte d'Ivoire	LK	Sri Lanka	SK	Slovakia
CM	Cameroon	LR	Liberia	SN	Senegal
CN	China	LT	Lithuania	SZ	Swaziland
CS	Czechoslovakia	LU	Luxembourg	TD	Chad
CZ	Czech Republic	LV	Latvia	TG	Togo
DE	Germany	MC	Monaco	TJ	Tajikistan
DK	Denmark	MD	Republic of Moldova	TT	Trinidad and Tobago
EE	Estonia	MG	Madagascar	UA	Ukraine
ES	Spain	ML	Mali	UG	Uganda
FI	Finland	MN	Mongolia	US	United States of America
FR	France	MR	Mauritania	UZ	Uzbekistan
GA	Gabon			VN	Viet Nam

METHOD OF TREATING MUSCULAR DISORDERS

The present invention relates to improvements in treating  
5 muscle disorders and related conditions using insulin-like  
growth factor I.

Insulin-like growth factor I (IGF-I) is a peptide present in  
the plasma and other body fluids. In its mature processed  
10 form it comprises 70 amino acids and can stimulate growth of  
a wide range of cell types. Human IGF-I have been cloned  
and its cDNA sequence can be found in Jansen et al, Nature,  
1993 (reference 11 below). The cDNA sequence encodes a  
precursor (also known as the D-chain), a mature 70 amino acid  
15 comprising the B, C and A regions respectively, and a C-  
terminal region which is termed the E-peptide. Recently, it  
has been found that the E-peptide can exist in different  
isoforms. This arises as a result of alternative splicing at  
the mRNA level. Chew et al report the existence of three  
20 alternatively spliced T-terminal regions of human IGF-I.  
One of the isoforms produced is as a result of splicing  
between exons 4, 5 and 6 of the gene and this predicts a  
prepro-IGF-I molecule of 158 amino acids including a C-  
terminal peptide, the Ec peptide of 24 amino acids in length.  
25 This Ec peptide appears to correspond to the Eb peptide  
found in rat IGF-I.

IGF-I has been proposed for use of a number of disorders  
relating to muscle atrophy and related conditions. For  
30 example, WO92/11865 proposes the use of human IGF-I for the  
prevention or treatment of cardiac disorders and for the  
promotion of cardiac muscle protein synthesis, for prevention  
or treatment of cardiomyopathies, acute heart failure or  
acute insult including myocarditis or myocardial infarction  
35 and for improving cardiac output by increasing heart/volume.  
WO95/13290 relates to the use of IGF-I for treating muscular

SUBSTITUTE SHEET (RULE 26)

- 2 -

disorders such as muscular dystrophy and related progressive skeletal muscle weakness and wasting.

WO93/09236 teaches methods of gene therapy using myogenic vector systems comprising promoters suitable for use in muscle cells. Such vectors may be introduced into a human patient for the treatment of muscle atrophy in ageing humans, muscle atrophy induced by spinal cord injuries or neuromuscular diseases.

10

A difficulty with the use of IGF-I is that this peptide is responsible for a wide range of effects within the human body. Although IGF-I is produced in muscle cells it is also produced in the liver from where it circulates and is involved in regulating metabolism. Administration of IGF-I may thus induce side-effects including hypoglycaemia.

De Vol et al (1990), Am. J. Physiol. 259, E89-E95) report that IGF-I expression is elevated during work-induced skeletal muscle growth. We have investigated the production of IGF-I in skeletal muscles and have surprisingly found that whereas resting muscles normally produce the liver IGF-I isoform including the Ea peptide, muscle cells induced to undergo rapid hypertrophy using active stretch rapidly up-regulate the production of a different IGF-I isoform. We have thus found that the IGF-I Ec isoform in humans, corresponding to the IGF-I Eb isoform in rats and rabbits, may play an important role in targeting the action of IGF-I to muscle cells. Thus, treatment of muscular disorders such as those mentioned above may be improved by the use of the human Ec isoform or the Ec peptide of human IGF-I.

Accordingly, the present invention provides a human IGF-I polypeptide or functional derivative thereof characterised by the presence of the Ec peptide for use in a method of treatment or therapy of the human or animal body. The

SUBSTITUTE SHEET (RULE 26)

- 3 -

invention also provides pharmaceutical compositions comprising such polypeptides.

5 The invention further provides a method for the treatment of muscular disorders of the human body which comprises administering to a patient in need of such treatment an effective amount of IGF-I or a functional derivative thereof which is characterised by the presence of Ec peptide.

10 The IGF-I isoform to which the invention relates may thus be used in methods of treating disorders relating to muscle atrophy and related conditions. This includes the use of human IGF-I for the prevention or cardiac disorders, diseases where promotion of cardiac muscle protein synthesis is a  
15 beneficial treatment, cardiomyopathies, acute heart failure or acute insult including myocarditis or myocardial infarction. The IGF isoform may also be used for improving cardiac output by increasing heart/volume.

20 Other muscular disorders which may be treated include muscular dystrophy, e.g. Duchenne or Becker muscular dystrophy, as well as autosomal dystrophies, and related progressive skeletal muscle weakness and wasting. The treatment of muscle atrophy in ageing humans, muscle atrophy  
25 induced by spinal cord injuries or neuromuscular diseases may also be treated by the present invention. Therapy with the appropriate IGF-I may also promote healing of bone fractures and maintenance of bone in old age.

30 Pharmaceutically acceptable carriers or diluents include those used in formulations suitable for oral or parenteral (e.g. intramuscular or intravenous) administration. The formulations may conveniently be presented in unit dosage form and may be prepared by any of the methods well known in  
35 the art of pharmacy. Such methods include the step of bringing into association the active ingredient with the

SUBSTITUTE SHEET (RULE 26)

- 4 -

carrier which constitutes one or more accessory ingredients. In general the formulations are prepared by uniformly and intimately bringing into association the active ingredient with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

For example, formulations suitable for parenteral administration include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents, and liposomes or other microparticulate systems which are designed to target the polypeptide to blood components or one or more organs.

The polypeptide of the invention may be administered by any suitable route, for example orally or injection, e.g. subcutaneous, intramuscular or intravenous injection, or alternatively may be produced in situ in a patient as a result of gene therapy treatment, for example as disclosed in WO93/09236, the contents of which are incorporated herein by reference.

WO95/13290, the contents of which are incorporated herein by reference, describes dose ranges of recombinant human IGF-I (the mature 70 amino acid polypeptide) in the range of 0.06 to 0.12 mg/kg per dose and IGF-I polypeptides comprising the Ec peptide may also be administered at this dose range. Doses may be administered at daily intervals or less frequently, for example at twice weekly or weekly intervals.

The IGF-I will preferably be a polypeptide comprising the 70 amino acid sequence of mature human IGF-I together with the Ec region which in humans corresponds to the Eb region of rabbit IGF-I set out in Figure 3 of the accompanying Example.

SUBSTITUTE SHEET (RULE 26)

- 5 -

The human Ec region is set out in Figure 4. Figure 5 shows the entire human cDNA sequence encoding the human Ea isoform.

While not wishing to be bound by any one particular theory we  
5 believe the human Ec isoform may be responsible for its  
activity via action on a receptor different from the normal  
IGF-I receptor. The presence of Ec region is believed to be  
responsible for binding to this receptor and thus derivatives  
of IGF-I Ec which retain the ability to induce the growth of  
10 muscle tissue may be used. Such derivatives may include  
further N-terminal truncations of the mature 70 amino acid  
sequence. This may be tested by bridging methods known as  
such to those of skill in the art, for example, by  
administration of such polypeptides (or mammalian equivalents  
15 thereof) to a mammal such as a rabbit or rat and observing  
the amount of muscle growth.

IGF-I Ec may be produced by any suitable means. Usually,  
this will be by recombinant means. For example, mRNA  
20 encoding the Ec isoform may be amplified using PCR primers as  
described in the accompanying Examples and the amplified  
product inserted into a suitable expression vector. IGF-I is  
produced by recombinant means commercially and these  
commercial methods may be used to produce the IGF-I Ec  
25 isoform. The vector may be any suitable recombinant vector  
known in the art for recombinant proteins. The vector will  
contain control signals for the expression of the IGF-I Ec  
protein operably linked to an open ring frame encoding said  
protein. The promoter will be compatible with a suitable  
30 host cell, for example a bacterial, yeast, insect or  
mammalian cell.

The following Example illustrates the invention.  
Insulin-like growth factor 1 (IGF-1) is a 70-residue  
35 polypeptide with important functions in the regulation of  
somatic growth, development and differentiation. The liver

SUBSTITUTE SHEET (RULE 26)

- 6 -

is a primary target for pituitary growth hormone (GH) which is stimulated to synthesize IGF-1. The resultant increase in circulating level of IGF-1 promotes cell division and is a major factor in regulating the growth of the body as a whole.

5 In several tissues there is also apparently a local system of growth regulation e.g. skeletal muscle which is able to undergo rapid hypertrophy to adapt to overload. It was, therefore, important to investigate the -role of IGF-1 in the locally regulated growth response.

10

Information on the organization of the IGF-1 gene is now available for several species, among which the human and rat genes are most extensively studied [1-5]. The structure of the IGF-1 gene is well conserved among mammals and contains  
15 several notable features, including an unexpectedly large size and the presence of alternatively spliced exons. In humans the IGF-1 gene comprises at least six exons (designated exon 1, 2, 3, 4, 5 and 6) spanning a region of over 90 kilobases (kb) of genomic DNA. Exons 1 and 2 are  
20 alternative leader exons [4,5] with distinct transcription start sites which are differentially spliced to the common exon 3 and produce class 1 and class 2 IGF-1 mRNA transcripts respectively [6-8]. Exons 3 and 4 code for the mature IGF-1 peptide (B, C, A and D domains) as well as the first 16 amino  
25 acid of the E Domain. Exons 5 and 6 each encodes an alternative part of a distinct extension peptide, named the E domain. This is followed by the termination codons of precursor IGF-1, 3' untranslated regions and poly(A) addition signal sites [2]. Sequence analyses of IGF-1 peptide  
30 purified from human plasma demonstrated that mature IGF-1 contains A, B, C and D domains. The A and B domains are homologous to the A and B chains of insulin [9].

Analysis of liver IGF-1 cDNA sequences also demonstrated the  
35 presence of an E peptide domain which was an extension of the D peptide domain [10-12]. A later study using antibodies

SUBSTITUTE SHEET (RULE 26)



- 7 -

directed against the E peptide of human IGF-1 confirmed that the mRNA sequence encoding the E peptide is actively translated and suggested that the E peptide circulates as part of the IGF-1 prohormone [13]. In rat liver IGF-1 mRNAs  
5 code for a 35-amino-acid E peptide sequence (IGF-1Ea). However an isoform (IGF-1Eb) with a different 41-amino-acid Eb domain has been detected at very low levels [14]. These two mRNAs encode alternative E peptide due to the presence (IGF-1Eb) or absence (IGF-1Ea) of a 52 base insert in the  
10 region coding the E domain [10, 14]. In human there are also IGF-1 cDNAs encoding three different Ea, Eb and Ec domains. The Ea and Eb-type cDNAs contain entirely different 3' sequences which specify different 3' untranslated sequences as well as different E domain coding sequences [2]. This is  
15 due to splicing in 3' exons [2]. The Ec is a exon 4-5-6 spliced cDNA which predicts a precursor IGF- 1 of 158 amino-acid residues and is the human counterpart of the rat Eb [15]. The physiological role of the alternative E peptide generated from IGF-1Ea, IGF-1Eb and IGF-1Ec remains unknown.  
20

Skeletal muscle has been shown to increase in mass very rapidly in response to passive stretch. The mature rabbit anterior tibialis is able to increase in mass by 35% in 4 days in this way [16]. From previous work [17, 18] this was  
25 known to be associated with the rapid production of new sarcomeres which are added serially at the ends of the fibre to existing myofibrils. Muscle stretch has been shown to result in an increase of IGF-1 mRNA as measured by RT-PCR [19]. However it is not know whether the IGF-1 gene is  
30 expressed by the muscle fibres themselves or by satellite cells or what isoforms of IGF-1 are involved. The regulation of muscle growth in vivo remains poorly understood, although the original observations on compensatory muscle hypertrophy implied that a component of muscle growth regulation is a  
35 localized, self-contained, and selflimiting process.

SUBSTITUTE SHEET (RULE 26)

- 8 -

Against this background, the current study was designed to determine whether local induction of muscle growth *in vivo* may involve alternative IGF-1 gene expression with different mRNA splicing for IGF-1 and localized fibre type expression.

## MATERIALS AND METHODS

1. **Animals and Muscle Stretch Procedure:** New Zealand white rabbits were used. The extensor digitorum. longus (EDL) muscle was subjected to acute stretch by immobilizing the left hind limb in, the extended position using a plaster cast. As was previously reported this results in a 35% increase in muscle mass within a few days [16]. After 6 days, euthanasia was induced by intravenous injection of an over dose of sodium pentobarbitone into the marginal ear vein. The EDL was immediately dissected out from both hind legs. The right hind leg served as the control. Each muscle was cut transversely into 2 parts, one part was fixed in freshly prepared 4% paraformaldehyde fixative at 4°C for 2 hours and later processed and embedded in paraffin wax. The second part was packed into a 1.5 ml tube and directly frozen in liquid nitrogen and stored at -70°C to await total RNA isolation.

2. **RNA isolation:** Total cellular RNA was isolated from stretched and normal muscle using the single-step method with acid guanidinium thiocyanate-phenol-chloroform extraction [20].

3. **Synthesis of probes for Northern blot and *in situ* hybridisation:** The oligonucleotide 5' TTGGGCATGTCAGTGTGG 3' which is complementary to the sequence of exon 4 of the IGF-1 gene was used as primer to synthesise cDNA of the IGF-1 mRNA by reverse transcriptase (RAV-2, Amersham). The cDNA was then amplified by PCR using two oligonucleotide primers (5'

SUBSTITUTE SHEET (RULE 26)

- 9 -

GCTTGCTCACCTTTACCAGC 3' and 5' TTGGGCATGTTCAGTGTGG 3'). A 280 base pair PCR product covering exon 3 and part of exon 4 of the IGF-1 gene was subcloned into pBS+ phagemid vector (Stratagene) including T3 and T7 promoters. Labelled sense and antisense RNA probes were synthesized by *in vitro* transcription with RNA polymerase using digoxigen in labelled uridine-triphosphate as substrate (Boehringer Mannheim) according to the manufacturers instructions. These probes were used for both Northern blotting and *in situ* hybridization.

4. **Northern blotting:** Samples containing the same amount (20  $\mu$ g) of total RNA were subjected to Northern blotting. The 280 bp antisense probe described above was used. Prehybridization (1 hour) and hybridization (15 hours) were carried out at 6°C in hybridization buffer [50% formamide; 5xSSC; 2% blocking reagent; 0.1% N-lauroylsarcosine; 0.02% SDS]. Washing was carried out at high stringency 2x5 minutes at room temperature with 1xSSC and 0.1% SDS, 2x 15 minutes at 68°C with 0.1xSSC and 0.1% SDS. The hybridized probe was detected by chemiluminescence according to the manufacturers instructions (Boehringer Mannheim). The blot filter was exposed to X-ray film for 6 hours.

5. ***In situ* hybridization:** The muscle tissues were cut in 10  $\mu$ m sections. Both transverse and longitudinal sections were taken and mounted onto autoclaved slides coated with 2% 3aminopropyltriethoxy-silane (Sigma). The sections were dewaxed by washing in xylene 3 times for 2 minutes each and rehydrated in a series of methanol solutions. The sections were then denatured by incubating in 0.2 N HCl at room temperature for 20 minutes, heated in 2xSSC at 70°C for 20 minutes and digested with pronase (10 $\mu$ g/ml, Boehringer Mannheim) in 50mM Tris.HCl for 15 minutes and was finally placed in 0.1 M triethanolamine (TEA) buffer, to which acetic anhydride was added to a final concentration of 0.5% and

SUBSTITUTE SHEET (RULE 26)

- 10 -

incubated for 10 minutes in order to block polar and charged groups in the sections. Hybridization was carried out in hybridization buffer [50% deionised formamide; 5 x SSC; 5 x Denhardt's solution; 250 µg/ml yeast t-RNA; 250 µg/ml denatured salmon sperm DNA; 4 mM ethylenediaminetetraacetic acid (EDTA) containing the DIG-labelled antisense or sense RNA probe consisting of the 280 bp sequence derived from exons 3 and part of exon 4 of the IGF-1 gene. The final concentration of probe was 1000 ng/ml. The hybridization was carried out at 68°C for 1 hour and then allowed to cool down to 42°C at which it was kept overnight in a humid chamber. After hybridization the sections were incubated with RNase A (100 µg/ml, Sigma) to remove the unbound single strand RNA probe. Washing was carried out at high stringency, 25 minutes in 2 x SSC at room temperature, 15 minutes in 1 x SSC at room temperature, 30 minutes in 0.5 x SSC at 42°C and 30 minutes in 0.5 x SSC at room temperature. The hybridized probe was detected by anti-digoxigenin-AP antibody conjugate, Fab fragments (1.5 U/ml, Boehringer Mannheim), according to manufacturers instructions.

6. Synthesis and molecular cloning of muscle IGF-1 cDNA: The first strand cDNA was synthesized by reverse transcriptase (RAV-2, Amersham.) from muscle total RNA with oligo dT primers and then amplified by the 3' rapid amplification of cDNA ends polymerase chain reaction (3' RACEPCR) with an IGF-1 gene specific primer (5' GCTTGCTCACCTTTACCAGC 3') which is part of the 5' end sequence of exon 3 of IGF-1. The PCR products were cloned into the pCR™ vector (Invitrogen) for DNA sequencing. Fragments were later sequenced by the dideoxy chain-termination method [2]. A total of 98% of the DNA sequence was obtained on both strands.

35

## RESULTS

SUBSTITUTE SHEET (RULE 26)

- 11 -

Northern Blotting. The results of Northern blot analysis performed with RNA extracted from normal and stretched extensor digitorum longus (EDL) are depicted in Fig. 1. The 280 bp IGF-1 antisense probe containing sequences derived from exon 3, and 4 of the IGF-1 gene hybridized with the two prominent IGF-1 mRNA species, 1.2 kb and 7.5 kb long. The expression of both types of mRNA species was greater in stretched muscle, although in some muscles the control muscle expressed more 7.5 kb mRNA than the stretched muscle.

10

**Localization of IGF-1 mRNA in normal and stretched muscle:** Expression of IGF-1 mRNA within normal and stretched muscle studied by *in situ* hybridization using antisense and sense RNA probe is shown in fig. 2. The *in situ* hybridization data demonstrates that the mRNA of IGF-1 is produced in response to stretch at the muscle fibre level as result of mechanical stimulation. This work showed that IGF-1 gene expression is not confined to the satellite cells but is up-regulated in the muscle fibres themselves. In transverse sections the IGF-1 message was localized to large muscle fiber but tended to be expressed strongly in the small fibres which represent the tapered ends of fibres terminating in the muscle belly [22]. In a few muscles some evidence of degeneration and regeneration was noted with high IGF-1 mRNA levels. These regions were superficial and indicated that in these cases the plaster cast was too tight. The *in situ* hybridization study however showed that with the use of the simple stretch model the upregulation of IGF-1 occurred in apparently undamaged fibres.

30

**Molecular cloning of muscle IGF-1 cDNAs:** This study was designed to investigate if different isoforms of IGF-1 are expressed in muscle when it is subjected to mechanical activity. Ten clones covering the E domain (exons three to six) were isolated and sequenced from stretched and from contralateral control muscle. Two classes of cDNA clone were

35

SUBSTITUTE SHEET (RULE 26)

- 12 -

obtained using RNA isolated from stretched muscle. Among these clones, 30% contain the sequences coding for IGF-1Ea and 70% for IGF-1Eb. However, even after repeated attempts, only IGF-1Ea type clones could be isolated from unstretched rabbit muscle. The cloned cDNA sequence starts from exon 3 which codes for mature IGF-1. The sequences of the two classes of IGF-1 cDNA isolated from total RNA of the stretched EDL muscle are shown in fig. 3. The sequence may be divided into three sections. A region which encodes mature IGF-1 (peptides B, C, A and D), an extension E peptide which in IGF-1Eb has a 52 base insert which is lacking in IGF-1Ea, and a common 3' untranslated region. In terms of the carboxyl-terminal extension (E) peptide, the rabbit amino acid sequence is identical to the human sequence up to residue E 16. At the first base of the codon for residue E 17, the amino acid sequences of the two cDNA clones diverge due to the 52-bp insert in the IGF-1Eb clone. The insert changes the derived amino acid sequence as well as the reading frame, resulting in two possible carboxyl-terminal E peptide sequences and the presence of two-different UAG stop codons in end variants.

Comparing the 52-bp insert from rabbit muscle with the 52-bp insert in the IGF-1Eb expressed in rat liver in very low amounts [14] and IGF-1Ec which has recently been detected in human liver [15], the positions where the insert occurs is the same. The rabbit cDNA sequence shows 77% homology with rat IGF-1Eb, with 12 of the 17 expected amino acid sequences being identical and 94% with human IGF-1Ec, with 13 of 16 expected amino acid sequence being identical (fig.4).

## DISCUSSION

Experimental models of muscle regeneration indicated that IGF-1 may act as a trophic factor in muscle regeneration [23] and it is expressed in proliferating myoblasts and satellite

SUBSTITUTE SHEET (RULE 26)

- 13 -

cells [24]. In this study we have analysed IGF-1 mRNA in skeletal muscle induced to undergo rapid longitudinal growth. This model was chosen as there is very little injury to the muscle fibres. The results presented here agree with published work [25-27] that IGF-1 mRNA is expressed in muscle tissues. However, they also show that the IGF-1 gene is expressed in the muscle fibres themselves and not solely in satellite and connective tissue cells. The expression of the IGF-1 transcripts was not uniform and it is usually the smaller fibres that show high levels of IGF-1 mRNA. A study of transverse and longitudinal sections showed that the small fibre which expressed IGF-1 mRNA also express the neonatal myosin heavy chain (MyHC). It has also been shown that small diameter fibres containing neonatal MyHC are the tapered ends of the larger fibres terminating within the belly of the muscle [22]. Longitudinal growth of skeletal muscle is facilitated by the addition of new sarcomeres to the ends of the existing myofibrils [28, 29] and the initial stage involves the laying down of neonatal myosin [22]. These data support the hypothesis that the ends of normal adult fibres are the region for longitudinal growth and that IGF-1 is involved in this process.

The estimation of the expression of the IGF-1 mRNA by Northern blotting suggests that both the 7.5 kb and 1.2 kb IGF-1 mRNA species are specifically induced by mechanical stimulation, but their increase seems to be independent of each other. The 1.2 kb mRNA was increased in all stretched muscle which was not always the case for the 7.5 kb mRNA. At this stage we do not know which is transcript for the IGF-1 Eb. Further work is needed to characterise these mRNA and to determine what coding signal sequence and other elements they include.

The isolation of two classes of cDNA clones (IGF-1Ea and IGF-1Eb) from stretched muscle indicates that both forms of IGF-

SUBSTITUTE SHEET (RULE 26)

- 14 -

I mRNA are present in the stretched muscle. The IGF-1Ea and IGF-1Eb cDNA 3' sequences differ by the presence of a 52-bp insert which in the latter alters the derived carboxyl-terminal amino acid sequence. Three mechanisms may account for the 52bp insert. Firstly, the insert could be generated by an alternate splice donor site 52-bp into the 5'-end of an intron present at this position in the IGF-1 genomic sequence. Alternatively, it may be generated by the use of an alternate splice acceptor site 52 bp from the 3' -end of the pertinent intron. Finally, the 52-bp insert could arise from a completely separate exon [10].

A comparison of the sequence of the rabbit IGF-1Eb with sequences of the rat IGF-1Eb [10] and human IGF-1Ec [15] showed that the IGF-1Eb which is markedly up-regulated in stretched muscle is apparently the rabbit counterpart to the rat IGF-1Eb and the human IGF-1Ec. Our results showed rabbit IGF-1Eb (the equivalent of human IGF-1Ec) was only detectable in stretched muscle. The fact that this is inducible isoform is consistent with the results of Chew et al [15] who demonstrated that after stimulation with physiological levels of GH, human IGF-1Ec transcript was increased in human hepatoma HepG2 cells (a hepatoma line), relative to human IGF-1Ea. The site for IGF-1 binding proteins (BP) is believed to be in the B domain [30]. Also C and D-domains, are thought to be "active regions" [31]. However, the physiological role of the alternative E peptide generated from IGF-1Ea and IGF-1Eb mRNA remains unknown [14]. It has been suggested that it could affect the interaction of IGF-1 with its receptor or its binding proteins. It has been also suggested that the E-peptides themselves may also have distinct biological roles after being cleaved from the prohormone [14]. Recently, part of the E-peptide has been shown to contain an amidated growth-promoting peptide with specific binding sites in human tissues [32]. According to our findings, the Eb-peptide appears to be induced only in

SUBSTITUTE SHEET (RULE 26)



- 15 -

the stretched muscle. This suggests the Eb-peptide may play a role in local growth control as exemplified by skeletal muscle fibre increasing in length and mass in response to mechanical stretch. The Eb peptide may be involved in the externalization of IGF-1 and also the binding of IGF-1 to muscle receptors.

There is evidence which suggests that the Ea peptide may be glycosylated in vivo. Bach et al [33] found that the Ea peptide can be glycosylated following in vitro translation in the presence of microsome. No putative glycosylation sites were noted from the muscle IGF-1 Eb sequence data. Possible functions for the differences in glycosylation of Ea and Eb include the reduction of the half life of IGF-1 1Eb, differential localization of the two forms and differential affinities for binding proteins. Therefore, the stretched muscle type IGF- 1Eb maybe much smaller but with a shorter half-life than the isoforms produced by normal muscle and the liver.

Devol et al [34] reported that IGF-1 mRNA in skeletal muscle is independent of GH and other pituitary hormones and demonstrated a link between local stimulation of skeletal muscle growth and IGF-1 gene expression. The Eb expressed only in stretched muscle indicates that the expression of IGF-1Eb mRNA might be switched on by mechanical stimulation, which is known to induce rapid muscle growth [16]. The Eb peptide may then be a specific factor distinguishing mechanical stimulation and associated mechanisms of muscle growth.

It is apparent from our study that the different E peptides may play different roles in IGF-1 activity. Further studies are required to elucidate whether the E peptide of these alternative IGF-1 mRNA interact differently with the IGF-1 receptor or with IGF-1 binding proteins, or whether

SUBSTITUTE SHEET (RULE 26)

- 16 -

the alternative E peptide alone enables the growth factor to act in an autocrine fashion.

## REFERENCES

- 5 [1] de Pagter-Holthuizen, P., van Schain, F.M.A, Verduijn G.M., van Ommen, G.J.B., Bouma, B.N., Jansen, M., and Sussenbach, J.S. (1986) FEBS Lett. 195, 179-184.
- [2] Rotwein, P., Pollock, KM., Didier, D.K, Krivi G.G.  
10 (1986) J. Biol. Chem 261,4828-4832.
- [3] Shimatsu, A. and Rotwein,P. (1987) J. Biol/ Chem 262, 7894-7900.
- [4] Tobin, G., Yee, D., Brunner, N. and Rotwein, P. (1990) Mol. Endocrinol. 4 (12), 1914-1920.
- 15 [5] Jansen, E., Steenbergh, P.H., LeRoith, D., Roberts, CT Jr. and Sussenbach J.S. (1991) Mol. Cell. Endocrinol. 78 (1-2), 115-125.
- [6] Adamo M.L., Ben-Hur, H., Roberts, CT Jr. and LeRoith, D. (1991) Mol. Endocrinol. 5(11), 1677-1686.
- 20 [7] Dickson, MC., Saunders JC. and Gilmour RS. (1991) J. Mol. Endocrinol. 6 (1), 17-31.
- [8] Weller, P.A., Dickson, M.C., Huskisson, N.S., Dauncey, M.J., Buttery P.J. and Gilmour, R.S. (1993) J. Mol. Endocrinol. 11, 201-211.
- 25 [9] Rinderknecht E. and Humbel R.E. (1987) J. Biol. Chem 253, 2769-2776.
- [10] Roberts, CT Jr., Lasky, S.T., Lowe, WL Jr. Seaman, W.T. and LeRoith, D. (1987) Mol. Endocrinol. 1, 243-248.
- [11] Jansen M. van Schaik F.M.A, Ricker A. T., Bullock B.,  
30 Woods, D.E., Gabbay K.R., Nussbaum, A.L., Sussenbach, J. S. and Van den Brande, J.L. (1983) Nature 306, 609-611.
- [12] Bell, G.I., Merryweather, J.P., Sanchez-Pescador, R., Stempien, M.M., Priestley, L., Scott, J. and Rall, LB. (1984) Nature 310, 775-777.
- 35 [13] Powell, D.R., Lee, P.D., Chang D. Liu, F. and Hintz, R.L. (1987) J. Clin. Endocrinol. Metab. 65, 868-875.

- 17 -

- [14] Lowe, WL Jr., Lasky, S.R., LeRoith, D. and Roberts CT Jr. (1988) Mol. Endocrinol. 2, 528-535.
- [15] Chew, S.L., Lavender P., Clark A.J.L. and Ross R.J.M. (1995) Endocrinology 136, 1939-1944.
- 5 [16] Goldspink, G., Scutt, A., Loughna P. T., Wells, D. J., Jaenicke, T. and Gerlach, G. F. (1992) Am. J. Physiol. 263 (3 Pt 2) R 356-363.
- [17] Tabary, J.C., Tabary, C., Tardieu C. Tardieu G. and Goldspink G. (1972) J. Physiol. 224(1), 231-244.
- 10 [18] Goldspink, D.F. Morton, A.J., Loughna, P. and Goldspink G. (1986) Pflugers Archiv-European Journal of Physiology 407 (3), 333-340.
- [19] Goldspink, D.F., Cox, V.M., Smith, S.K, Eaves, L.A., Osbaldeston, N.J., Lee, D.M. and Mantle, D. (1995) Am. J. Physiol. 268, E288-E297.
- 15 [20] Chomczynski, P. and Sacchi, N, (1987) Analyt. Biochem. 162, 156-159.
- [21] Sanger, F., Nicklen, S. and Coulson, A.R. (1977) Proc. Natl. Acad. Sci. USA 74, 5463-5467.
- 20 [22] Benjamin, W.C., Donna, M.W., Stacey, D.L., Jacalyn, D.A. and Everett, B. (1995) The Anatomical Record 242, 462-470.
- [23] Jennische, E., Skottner, A. and Hansson, H.A. (1987) Acta Physiol. Scand. 129, 9.
- 25 [24] Edwall, D., Schalling, M., Jennische, E. and Norstedt, G. (1989) Endocrinology 124, 820-825.
- [25] Beck, F., Samani, N.J., Penschow, J.D., Thorley, B., Tregear, G.W. and Coghlan, J.P. (1987) Development 101, 175-184.
- 30 [26] Han, V.K.M., D'Erocole, A.J. and Lund, P.K (1987) Science 236, 193-197.
- [27] Caroni, P. and Schneider C. (1994) J. Neurosci. 14, 3378-3388.
- [28] Williams, P.E. and Goldspink, G. (1971) J. Cell Sci 9, 751-767.
- 35 [29] Williams, P.E. and Goldspink, G. (1973) J. Anat.

SUBSTITUTE SHEET (RULE 26)

116,45-46.

[30] DeVroede, M.A., Rechler, M.M., Nissley, S.P., Josni, S., Burke, G.T. and Katsoyannis, P.G. (1985) Proc. Natl. Acad. Sci. USA 82, 3010-3014.

- 5 [31] Pietrkowski, Z., Wernicke, D., Porcu, P., Jameson, B.A. and Baserga, R. (1992) Cancer Res. 52, 6447-6451.

[32] Siegfried, L.M., Kasprzyk, P.G., Treston, A.M., Mulshine J. L., Quinn, KA and Cuttitta F. (1992) Proc. Natl. Acad. Sci. USA 89 (17), 8107-8111.

- 10 [33] Bach M.A., Roberts CT Jr., Smith E.P. and LeRoith, D. (1990) Mol Endocrinol. 4(6), 899-904.

[34] Devol, D.L., Rotwein, P., Sadow, J.L., Novakofski, J. and Bechtel, P.J. (1990) Am. J. Physiol. 259, E89-E95.

# 15 Figure Legends

**Fig. 1.** Expression of IGF-1 mRNA studied by Northern blotting in stretched (E) and control (C) extensor digitorum longus (EDL) muscle.

- 20 **Fig. 2.** Localization and distribution of IGF-1 mRNA in stretched (A, transvers section; B, longitudinal section) and control (C) extensor digitorum longus (EDL) muscle. The sense RNA probe from the same clone was used on the stretched muscle (D) as a negative control. Scale bar, 30  
25  $\mu$ m.

- Fig. 3.** DNA and derived amino acid sequences of rabbit IGF-1 cDNA isolated from stretched muscle: The two types of cDNA sequence differ by the presence (IGF-1Eb) or absence (IGF-1Ea) of a 52 base pair insert (underline) from position  
30 288 through position 340. The insert altered the derived C-terminal amino acid sequence of the E peptide (underline in IGF-1Eb case), changed the reading frames and used two different TAG stop codons (end). The putative glycosylation site (Asn-Thr-Ser) (marked by ♦♦♦) is present  
35 in the Ea but not in the Eb peptide.

**Fig. 4.** Alignment of the three derived amino acid sequences

- 19 -

of the inserts from rat liver (bottom), human liver (middle) and rabbit stretched muscle (top). Identical amino residues are shown by the boxes.

SUBSTITUTE SHEET (RULE 26)

- 20 -

CLAIMS

1. An IGF-I polypeptide or functional derivative thereof characterised by the presence of the Ec peptide or  
5 functional equivalent thereof for use in a method of treatment or therapy of the human or animal body.
2. A human IGF-I polypeptide or functional derivative thereof characterised by the presence of the Ec peptide for use in a method of treatment or therapy of the  
10 human body.
3. An animal IGF-I polypeptide or functional derivative thereof characterised by the presence of the Ec peptide or functional equivalent thereof for use in a method of treatment or therapy of the animal body.
- 15 4. A pharmaceutical composition comprising a human IGF-I polypeptide or functional derivative thereof characterised by the presence of the human Ec peptide, together with a carrier or diluent.
5. A method for the treatment of muscular disorders  
20 of the human or animal body which comprises administering to a patient in need of such treatment and which is characterised by the presence of Ec peptide or functional equivalent thereof.
6. A method for the treatment of muscular disorders  
25 of the human body which comprises administering to a human patient in need of such treatment an effective amount of human IGF-I or a functional derivative thereof which is characterised by the presence of Ec peptide.
7. A method for the treatment of muscular disorders  
30 of the animal body which comprises administering to an animal patient in need of such treatment an effective amount of animal IGF-I or a functional derivative thereof which is characterised by the presence of Ec peptide or functional equivalent thereof.
- 35 8. In a method of treating a muscular disorder of the human or animal body which comprises administering to a

SUBSTITUTE SHEET (RULE 26)

- 21 -

patient in need of such treatment an effective amount of IGF-I, in the improvement comprising administration of IGF-I or a functional derivative thereof which is characterised by the presence of Ec peptide or functional equivalent thereof.

9. In a method of treating a muscular disorder of the human body which comprises administering to a human patient in need of such treatment an effective amount of human IGF-I, the improvement comprising administration of IGF-I or a functional derivative thereof which is characterised by the presence of Ec peptide.

10. In a method of treating a muscular disorder of the animal body which comprises administering to an animal patient in need of such treatment an effective amount of animal IGF-I, the improvement comprising administration of animal IGF-I or a functional derivative thereof which is characterised by the presence of EC peptide or functional equivalent thereof.

11. An IGF-I polypeptide or functional derivative thereof for use according to any one of claims 1 to 3, or a method according to any one of claims 5 to 10 wherein the muscular disorder is muscular dystrophy.

SUBSTITUTE SHEET (RULE 26)

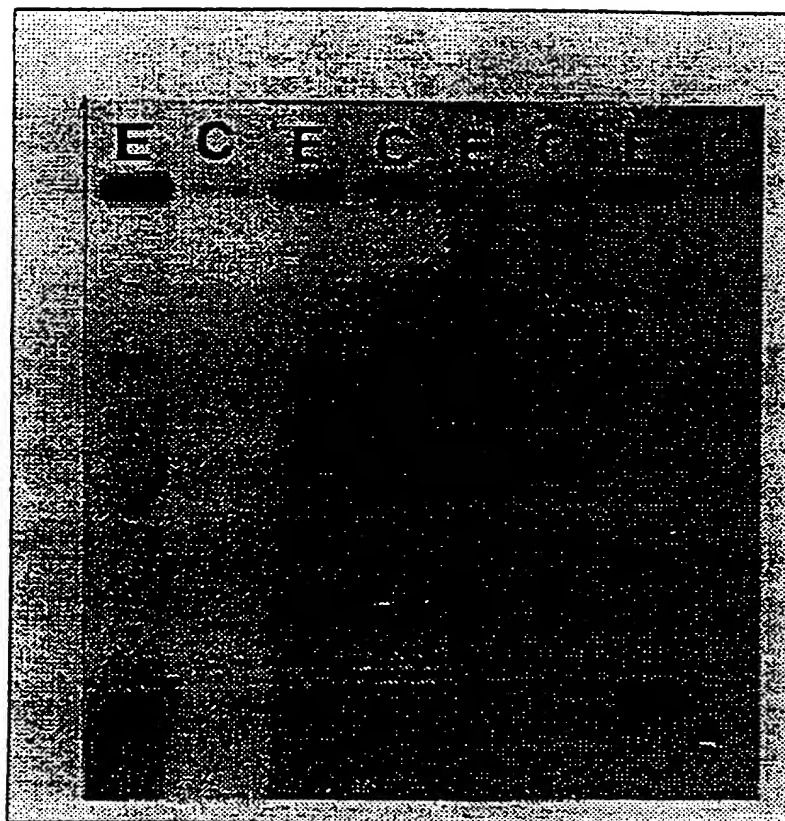


FIG. 1

SUBSTITUTE SHEET (RULE 26)



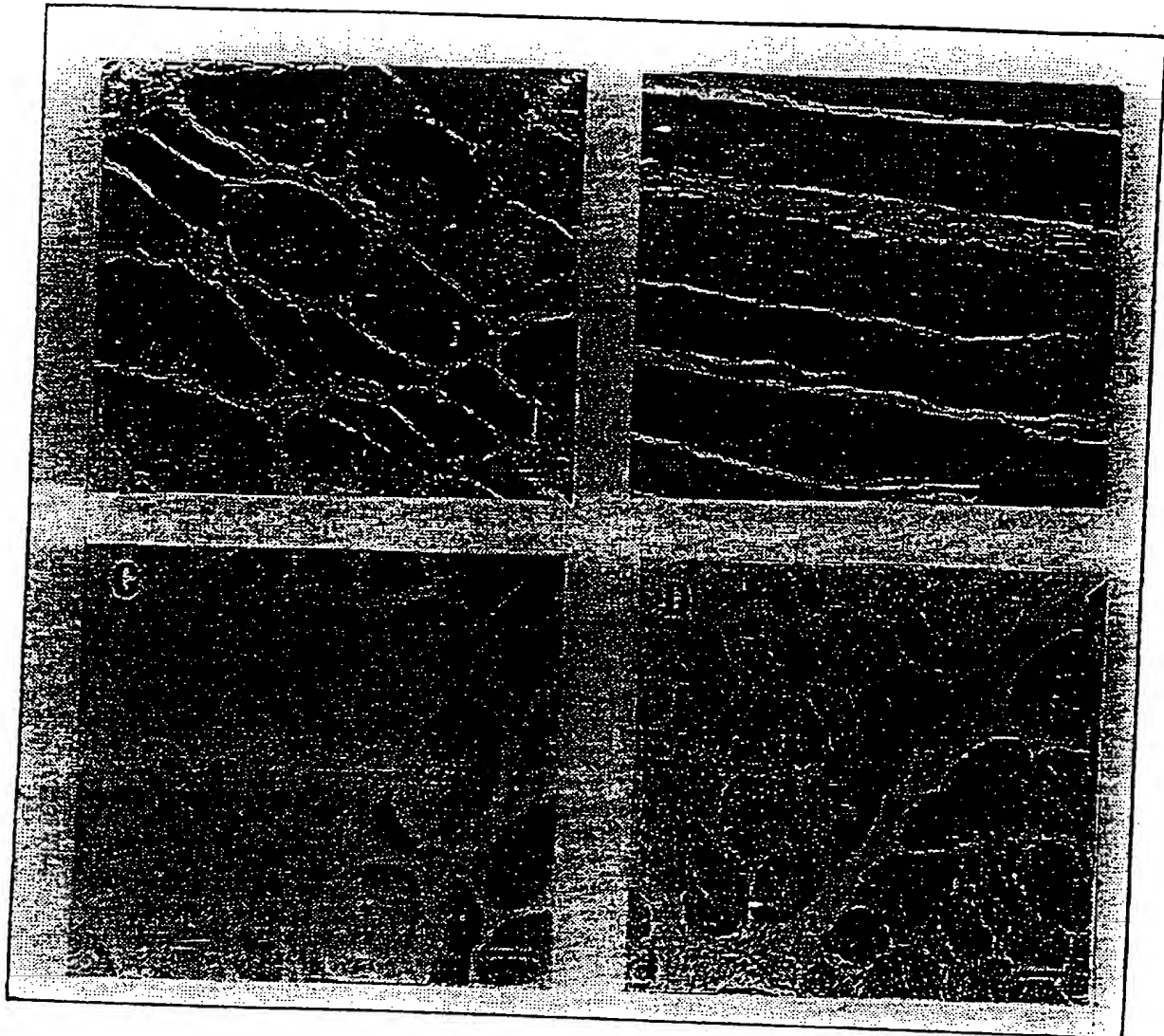


FIG. 2

3/6

TTGCTCACCTTTACCAGCTCGGCCACAGCCGGACCGGAGACGCTCTGCGGTGCTGAGCTG 60  
 LeuLeuThrPheThrSerSerAlaThrAlaGlyProGluThrLeuCysGlyAlaGluLeu  
 B-->

GTGGATGCTCTTCAGTTCGTGTGTGGAGACAGGGGCTTTTATTTCAACAAGCCACAGGA 120  
 ValAspAlaLeuGlnPheValCysGlyAspArgGlyPheTyrPheAsnLysProThrGly  
 C->

TACGGCTCCAGCAGTCCGAGGGCACCTCAGACAGGCATCGTGGATGAGTGCTGCTTCCGG 180  
 TyrGlySerSerSerArgArgAlaProGlnThrGlyIleValAspGluCysCysPheArg  
 A-->

AGCTGTGATCTGAGGAGGCTGGAGATGTACTGTGCACCCCTCAAGCCGGCAAAGGCAGCC 240  
 SerCysAspLeuArgArgLeuGluMETTyrCysAlaProLeuLysProAlaLysAlaAla  
 D-->

288  
 CGCTCCGTCCTGCCCCAGCGCCACACCGACATGCCCAAGACTCAGAAGTATCAGCCTCCA 300  
 ArgSerValArgAlaGlnArgHisThrAspMETProLysThrGlnLysTyrGlnProPro  
 E--> Eb-->

340  
 TCTACCAACAAGAAAATGAAGTCTCAGAGGAGAAGGAAAGGAAGTACATTTGAAGAACAC 360  
 SerThrAsnLysLysMETLysSerGlnArgArgArgLysGlySerThrPheGluGluHis  
 GluValHisLeuLysAsnTh  
 Ea--> ◆ ◆

AAGTAGAGGGAGTGCAGGAAACAAGAACTACAGGATGTAGGAAGACCCTTCTGAGGAGTG 420  
 Lysend  
 rSerArgGlySerAlaGlyAsnLysAsnTyrArgMETend

◆  
 AAGAAGGACAGGCCACCGCAGGACCCTTTGCTCTGCACAGTTACCTGTAAACATTGGAAT 480  
 ACCGGCCAAAAATAAGTTTGATCACATTTCAAAGATGGCATTTCCTCCCAATGAAATACA 560  
 CAAGTAAACATTC

FIG. 3

SUBSTITUTE SHEET (RULE 26)

4/6

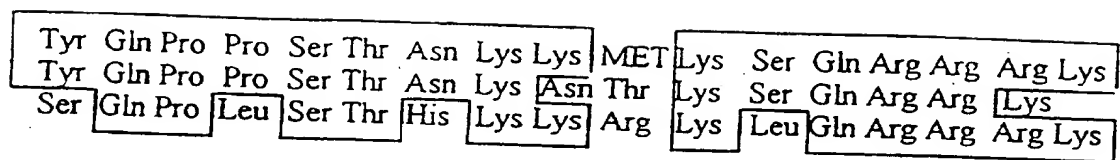


FIG. 4

SUBSTITUTE SHEET (RULE 26)

5/6

## HUMAN IGF-I Ea isoform

## SIGNATURE

Pst-I  
 5'-CTGCAGGGGGGGGGGGGGGGGGCTTCA GAA GCA ATG GGA AAA ATC AGC AGT 50  
 Ser Glu Ala Met Gly Lys Ile Ser Ser  
 -50

CTT CCA ACC CAA TTA TTT AAG TGC TGC TTT TGT GAT TTC TTG AAG GTG 100  
 Leu Pro Thr Gln Leu Phe Lys Cys Cys Phe Cys Asp Phe Leu Lys Val  
 -40 -30

AAG ATG CAC ACC ATG TCC TCC TCG CAT CTC TTC TAC CTG GCG CTG TGC  
 Lys Met His Thr Met Ser Ser Ser His Leu Phe Tyr Leu Ala Leu Cys  
 -20

150  
 CTG CTC ACC TCC ACC AGC TCT GCC ACG GCT GGA CCG GAG ACG CTC TGC  
 Leu Leu Thr Ser Thr Ser Ser Ala Thr Ala Gly Pro Glu Thr Leu Cys  
 -10 -1 1

200  
GGG GCT GAG CTG GTG GAT GCT CTT CAG TTC GTG TGT GGA GAC AGG GGC  
 Gly Ala Glu Leu Val Asp Ala Leu Gln Phe Val Cys Gly Asp Arg Gly  
 10 20

250  
TTT TAT TTC AAC AAG CCC ACA GGG TAT GGC TCC AGC AGT CGG AGG GCG  
 Phe Tyr Phe Asn Lys Pro Thr Gly Tyr Gly Ser Ser Ser Arg Arg Ala  
 30

300  
CCT CAG ACA GGT ATC GTG GAT GAG TGC TGC TTC CGG AGC TGT GAT CTA  
 Pro Gln Thr Gly Ile Val Asp Glu Cys Cys Phe Arg Ser Cys Asp Leu  
 40 50

Fig 5

SUBSTITUTE SHEET (RULE 26)

6/6

350  
 AGG AGG CTG GAG ATG TAT TGC GCA CCC CTC AAG CCT GCC AAG TCA GCT  
 Arg Arg Leu Glu Met Tyr Cys Ala Pro Leu Lys Pro Ala Lys Ser Ala  
 60 70

400  
 CGC TCT GTC CGT GCC CAG CGC CAC ACC GAC ATG CCC AAG ACC CAG AAG  
 Arg Ser Val Arg Ala Gln Arg His Thr Asp Met Pro Lys Thr Gln Lys  
 80

450  
 GAA GTA CAT TTG AAG AAC GCA AGT AGA GGG AGT GCA GGA AAC AAG AAC  
 Glu Val His Leu Lys Asn Ala Ser Arg Gly Ser Ala Gly Asn Lys Asn  
 90 100

500  
 TAC AGG ATG TAG GAAGACCCTCCTGAGGAGTGAAGAGTGACATGCCACC  
 Tyr Arg Met \*\*\*

550  
 GCAGGATCCTTTGCTCTGCACGAGTTACCTGTAACTTTGGAACACCTACCAAAAAATA

600 650  
 AGTTTGATAACATTTAAAAGATGGGCGTTTCCCCCAATGAAATACACAAGTAAACATTCC

700  
 AACATTGTCTTTAGGAGTGATTTGCACCTTGCAAAAATGGTCCTGGAGTTGGTAGATTGC

750  
 TGTTGATCTTTTATCAATAATGTTCTATAGAAAAGAAAAAAAAACCCCCCCCCCCCCCT

GCAGT-3'  
 Pst-I

Fig 5 (continued)

SUBSTITUTE SHEET (RULE 26)

## INTERNATIONAL SEARCH REPORT

Inter national Application No  
PCT/GB 97/00658A. CLASSIFICATION OF SUBJECT MATTER  
IPC 6 C12N15/16 C07K14/65 A61K38/30

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 229 750 A (UNIV WASHINGTON) 22 July 1987 see the whole document, especially page 6, lines 5-8 and Figs. 2 and 6 ---	1-4,11
Y	WO 95 13290 A (NORTH SHORE UNIVERSITY HOSPITAL) 18 May 1995 cited in the application see the whole document ---	1-11
Y	SCIENCE, vol. 236, 1987, pages 193-197, XP000676548 HAN V.K. ET AL.: "Cellular localization of somatomedin (insulin-like growth factor) messenger RNA in the human fetus." cited in the application see the whole document, especially Table 1 --- -/-	1-11

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*A\* document member of the same patent family

Date of the actual completion of the international search

30 June 1997

Date of mailing of the international search report

11.07.97

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,  
Fax (+ 31-70) 340-3016

Authorized officer

Mandl, B

Intern      Application No  
PCT/GB 97/00658

Intern      Application No  
PCT/GB 97/00658

Form PCT/ISA/210 (continuation of second sheet) (July 1992)

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/GB 97/00658

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>JOURNAL OF PHYSIOLOGY, vol. 495P, 2 July 1996, pages 162p-163p, XP000677210 GOLDSPINK G. ET AL.: "Local growth regulation is associated with an isoform of IGF-1 that is expressed in normal muscle but not in dystrophic mdx or dydy mouse muscles when subjected to stretch." see abstract</p>	1-11
T	<p>--- JOURNAL OF MUSCLE RESEARCH AND CELL MOTILITY, vol. 17, August 1996, pages 487-495, XP000677224 YANG S. ET AL.: "Cloning and characterization of an IGF-1 isoform expressed in skeletal muscle subjected to stretch." see the whole document -----</p>	1-11



# INTERNATIONAL SEARCH REPORT

International application No.

PCT/GB 97/ 00658

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 5-10  
because they relate to subject matter not required to be searched by this Authority, namely:  
Remark: Although claims 5-10 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. ☐ Claims Nos.:  
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/GB 97/00658

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0229750 A	22-07-87	US 4963665 A	16-10-90
		AU 588072 B	07-09-89
		AU 6715187 A	09-07-87
		DE 3789559 D	19-05-94
		DE 3789559 T	13-10-94
		ES 2062993 T	01-01-95
		IL 81175 A	26-08-94
		JP 8019158 B	28-02-96
		JP 62228285 A	07-10-87
		US 5070075 A	03-12-91
-----			
WO 9513290 A	18-05-95	AU 1178395 A	29-05-95
-----			